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STEREOCONTROLLED SYNTHESIS OF DITHYMIDINE PHOSPHOROTHIOATES BY USE OF A FUNCTIONALIZED 5'-PROTECTING GROUP BEARING AN IMIDAZOLE RESIDUE[†]

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ABSTRACT: Diastereoselective formation of internucleotidic phosphorothioate triester bonds was achieved by use of 3-(imidazol-1-ylmethyl)-4',4"-dimethoxytrityl (IDTr) as a 5'-hydroxyl protecting group in the phosphotriester approach. After removal of all the protecting groups, stereochemistry of the major product was determined as the Sp-configuration by enzymatic digestion.

Phosphorothioate analogs of oligodeoxyribonucleotides have been widely used as reliable antisense DNAs for selective inhibition of gene expression. Phosphorothioate DNAs can be routinely synthesized by the automated solid-phase phosphoramidite and H-phosphonate method with sulfurization of the phosphite and H-phosphonate intermediates, respectively. However, in both methods a random mixture of Sp and Rp isomers is formed. It has been demonstrated that all-Sp phosphorothioates have higher affinity for their complementary oligonucleotides than all-Rp diastereoisomers. Therefore, there is much current interest in developing improved methods for the stereocontrolled synthesis of phosphorothioate DNAs.

On the other hand, we have reported that the use of 3-(imidazol-1-ylmethyl)-4',4"-dimethoxytrityl (IDTr) as a 5'-hydroxyl protecting group resulted in remarkable enhancement of the rate of internucleotidic bond formation.⁶ Recently, it was found that the IDTr group-catalyzed condensation of 5'-hydroxyl components with P-prochiral S-phenyl deoxynucleoside 3'-phosphorothioates proceeded in a stereospecific manner to some extent.⁷

[†] This paper is dedicated to the memory of the late Professor Tsujiaki Hata.

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In this paper, we wish to describe a new approach to stereocontrolled formation of internucleotidic phosphorothicate triester bonds by use of the IDTr group as the 5'-hydroxyl protecting group in the phosphotriester approach.

RESULTS AND DISCUSSION

In a previous paper, we demonstrated that condensation of S-phenyl 5'-O-IDTr-thymidine 3'-phosphorothioate (4a) with 3'-O-benzoylthymidine (7) in the presence of isodurenedisulfonyl dichloride (DDS)⁸ resulted in predominant formation of one of the diastereomers of the product 8a in the ratio of 85:15.⁷ In extension of our study, substituent effects of the S-phenyl group were examined. Phosphorylating reagents 2b-d were synthesized in 56-86% yields by a procedure similar to that described in the synthesis of 2a.⁹ By use of these reagents, S,S-diaryl 5'-O-IDTr-thymidine 3'-phosphorodithioates 3c and 3d were synthesized in 90% and 93% yields, respectively. The resulting compounds were treated with H₂O-Et₃N-pyridine (1:2:2, v/v/v) to give the corresponding phosphodiester components 4c and 4d quantitatively. In the case of 3b, one of the 2,6-dichlorophenylthio groups was lost during the aqueous workup, and 4b obtained directly in 65% yield.

Next, we examined the condensation of 4b-d with 1 equiv of 3'-Obenzoylthymidine (7) in the presence of 3 equiv of DDS as a condensing reagent. TLC monitoring of the reaction mixture indicated that the reactions were completed within 10 min in all cases and almost quantitative formation of the dimers 8b-d was observed. After extraction, the crude products were dissolved in CDCl₃ and the ratios of the diastereomers were estimated by ³¹P NMR analysis. The results are listed in TABLE 1. FIG. 1 shows the ³¹P NMR spectrum of 8c. The ratio of the diastereomers which appeared at 24.11 and 24.40 ppm is estimated to be 86:14. In the case of 8b, partial elimination of the 2,6-dichlorophenylthio group from the product was observed during the aqueous treatment. Therefore, the diastereomeric ratio of the dimer 8b did not show an accurate value for the internucleotidic bond formation. In spite of the substitution of the phenylthio group by larger arylthio groups, no significant difference in the ratio of the diastereomers was observed in these reactions. As a reference reaction, when Sphenyl 5'-O-DMTr-thymidine 3'-phosphorothioate was used in place of 4a in the presence of 1 equiv of N-methylimidazole and 3 equiv of DDS, a 1:1 mixture of the corresponding diastereomers was obtained. These results suggested that the predominant formation of one of the diastereomers is apparently attributed to the intramolecular assistant of the imidazole residue as shown in SCHEME 1. When 4a was activated with an excess amount of DDS in the absence of the 5'-hydroxyl component, formation of the relatively stable phosphorochloridate intermediate 5a was observed as a nearly 1:1

TABLE 1. Condensation of S-aryl 5'-O-IDTr-thymidine 3'-phosphorothioates **4a-d** with **7** in the presence of DDS.

	ArS	monomer unit 4 ³¹ P NMR (CDCI ₃)	dimer 8 ³¹ P NMR (CDCI ₃)	ratio of diastereomers
а	s	12.52	24.77, 25.09	85 : 15
b	CI CI	11.97	21.11, 21.51	77 : 23
c		13.75	24.11, 24.40	86 : 14
d	S s	13.77	23.99, 24.41	86 : 14

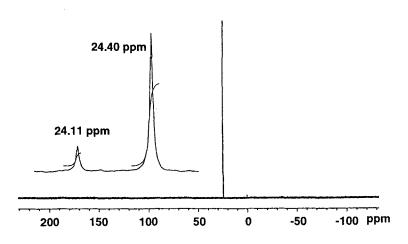


FIG. 1. ³¹P NMR spectrum of 8c in CDCl₃ after aqueous workup.

SCHEME 1

mixture of the diastereomers by ³¹P NMR spectroscopy.^{6,10} According to the remarkable acceleration of the rate of condensation and enrichment of the stereoselectivity caused by the IDTr group, the existence of a cyclic intermediate **6** is highly plausible. It is likely that steric bulk resulted from formation of an intramolecular cyclic structure restricts the direction of the nucleophilic attack of the nucleophile at the phosphorus center.

These results prompted us to study stereocontrolled synthesis of dinucleoside phosphorothioate diesters. The IDTr-assisted condensation system requires neither optically pure starting materials nor optically pure reactive intermediates. Therefore, we synthesized racemic 5'-O-IDTr-thymidine 3'-O-(2-chlorophenyl)phosphorothioate (9) as the starting material. 5'-O-IDTr-thymidine (1) was allowed to react with in situ prepared (2-chlorophenyl)phosphorobis(1,2,4-triazolide)¹¹ in THF. The resulting phosphorotriazolide intermediate was further treated with hexamethyldisilathiane as an anhydrous H₂S equivalent. ^{12,13} After the usual workup, a diastereomixture of **9** (53.51, 54.27 ppm) was obtained in 58% yield. Next, condensation of 9 with 1 equiv of the 5'-hydroxyl component (7) in the presence of 3 equiv of DDS was elucidated by means of ³¹P NMR (FIG. 2). After 1 h, the yield of the dimer 12 (62.66, 63.11 ppm) was estimated to be 92% and the ratio of the diastereomers was 18:82. It was noteworthy that formation of the phosphotriester resulting from S-activation was less than 1%. Activation of 9 with 3 equiv of DDS in the absence of 7 gave a nearly 1:1 diastereomeric mixture of the phosphorochloridate intermediates 10 (62.70, 63.00 pm). In addition, small signals of pyrophosphorothioate derivatives (less than 5%) were observed in the region around 45 to 50 ppm.

The diastereomeric mixture of 12 was treated with 1% TFA followed by tetramethylguanidinium 2-pyridinaldoximate to remove the 5'-O-IDTr and 2-chlorophenyl groups. Finally, the 3'-benzoyl group was removed by aqueous ammonia treatment and the fully deprotected dinucleoside phosphorothioate 13 was obtained. After purification by paper chromatography, dithymidine phosphorothioate 13 was obtained in 42% yield. Reversed phase HPLC analysis of the purified product indicated that the ratio of the diastereomers was 23:77. Difference in the diastereomeric ratio between 12 and 13 might be due to the racemization of 12 during the oximate treatment or a partial loss of the one of the diastereomers of 13 during purification. The major isomer was separated by HPLC and treated with snake venom phosphodiesterase and nuclease P1. The major isomer was resistant to snake venom phosphodiesterase (FIG. 3B) and was completely digested by nuclease P1 (FIG. 3C). These results clearly indicated the major product had

SCHEME 2

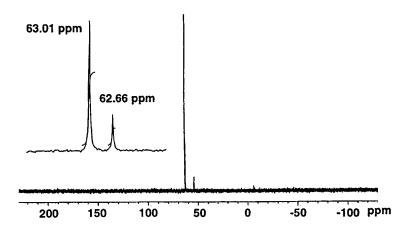


FIG. 2. ^{31}P NMR spectrum of the mixture obtained by the reaction of 9 with 7 in the presence of DDS in pyridine- C_5D_5N (9:1, v/v).

SCHEME 3

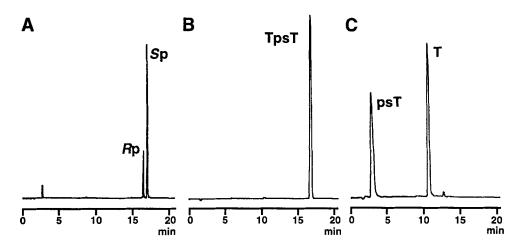


FIG. 3. (A) Reversed phase HPLC profile of the diastereomeric mixture of TpsT. (B) Snake venom phosphodiester digestion of the major isomer. (C) Nuclease P1 digestion of the major isomer.

the Sp configuration.^{14,15} During removal of the phosphate protecting group of 12, attack of the oximate anion at the phosphorus center caused inversion of the configuration. Therefore, absolute configuration of the major isomer of 12 was determined to be Rp.

In conclusion, the present IDTr-catalyzed condensation enabled us to synthesize dinucleoside phosphorothicate bearing the Sp configuration as the major product. The method requires neither diasteromerically pure starting materials nor reactive intermediates.

EXPERIMENTAL

General. Pyridine was distilled after being refluxed over *p*-toluenesulfonyl chloride for several hours, redistilled from CaH₂, and stored over molecular sieves 4A. ¹H NMR spectra were obtained at 270 MHz on a JEOL-EX-270 spectrometer with tetramethylsilane as an internal standard in CDCl₃. ¹³C NMR spectra were obtained at 67.8 MHz on a JEOL-EX-270 spectrometer with tetramethylsilane as an internal standard. ³¹P NMR spectra were obtained at 109.25 MHz on a JEOL-EX-270 spectrometer using 85% H₃PO₄ as an external standard. UV spectra were recorded on a Hitachi 220A spectrophotometer. Thin layer chromatography was performed on precoated glass plates of Kieselgel 60 F₂₅₄ (Merck, No. 5715). Silica gel column chromatography was carried out using Wakogel C-200. Reversed phase HPLC was performed on a column of μBondasphere 5-μm C18 100 Å, 3.9 mm x 15 cm (Nihon Waters Ltd.) with a linear gradient of 0-60% acetonitrile in 0.1 M ammonium acetate buffer (pH 7.0) at 50 °C for 60 min at a rate of 1.0 mL/min.

Cyclohexylammonium S,S'-Bis(2,6-dichlorophenyl) Phosphorodithioate (2b). To dry pyridine (16 mL) cooled at 0 °C was added dropwise methyl phosphorodichloridate (1.11 mL, 11.2 mmol) under argon. The mixture was stirred at 0 °C for 15 min. A mixture of 2,6-dichlorobenzenethiol (4.0 g, 22.3 mmol) and triethylamine (3.11 mL, 22.3 mmol) in dry pyridine (5 mL) was added dropwise to the above solution. After being stirred at rt for 3 h, the mixture was treated with water (20 mL), diluted with CHCl₃, and transferred into a separating funnel. The organic layer was washed three times with water, and the aqueous layer was back-extracted three times with CHCl3. The organic layer and washings were combined, dried over Na2SO4, filtered, and concentrated under reduced pressure. The residue was coeyaporated three times with toluene and dissolved in CHCl₃ (20 mL). Cyclohexylamine (2.55 mL, 22.3 mmol) was added dropwise to the above solution. After the mixture was stirred at rt for 12 h, the resulting white solid was collected by filtration and washed with ether. Recrystallization of the white powder from ethanol gave 2b (4.83 g, 83%) as a white needle: mp 185-186 °C; 31 P NMR (C₅D₅N) δ 19.64; 1 H NMR (C₅D₅N) δ 1.17-1.85 (10H, m, 2,3,4,5,6-H of cyclohexylamine), 2.83 (1H, m, 1-H of cyclohexylamine), 7.25 (2H, t, J = 6.9 Hz, 4-H of dichlorophenyl), 7.42 (4H, d J = 6.9 Hz, 3.5-H of)dichlorophenyl), 7.79 (2H, bs, NH₂); ¹³C NMR (C₅D₅N) δ 23.89, 24.53, 49.49, 128.18, 129.00, 129.54, 129.58, 132.20, 132.31, 141.04, 141.12. Anal. Calcd for C₁₈H₂₀Cl₄NO₂PS₂: C, 41.63; H, 3.88; N, 2.70; S, 12.35. Found: C, 41.22; H, 3.95; N, 2.62; S, 12.52.

Cyclohexylammonium S,S'-Bis(1-naphthyl) Phosphorodithioate (2c). The same procedure as described in the synthesis of 2b by use of 1-naphthalenethiol (2.12 g, 13.2

mmol) gave 2c (1.78 g, 56%) as a white needle: mp 184-187 °C; ³¹P NMR (C₅D₅N) δ 27.56; ¹H NMR (C₅D₅N) δ 0.92-2.07 (10H, m, 2,3,4,5,6-H of cyclohexylamine), 2.91 (1H, m, 1-H of cyclohexylamine), 7.31-7.45 (6H, 3,6,7-H of 1-naphthyl), 7.85 (4H, t, J = 14.4 Hz, 5,8-H of 1-naphthyl), 8.58 (2H, d, J = 5.4 Hz, 2-H of 1-naphthyl), 8.68 (2H, d, J = 5.4 Hz, 4-H of 1-naphthyl), 9.36 (2H, bs, NH₂); ¹³C NMR (C₅D₅N) δ 22.23, 22.62, 28.84, 47.85, 123.65, 123.77, 123.94, 124.60, 125.95, 125.98, 126.13, 129.04, 129.13, 131.21, 131.29, 132.11, 132,49. Anal. Calcd for C₂₆H₂₈NO₂PS₂: C, 64.84; H, 5.86; N, 2.91; S, 13.32. Found: C, 65.43; H, 5.85; N, 2.65; S, 13.15.

Cyclohexylammonium *S,S'*-**Bis**(**2-naphthyl**) **Phophorodithioate** (**2d**). The same procedure as described in the synthesis of **2b** by use of 2-naphthalenethiol (10.76 g, 67.2 mmol) gave **2d** (11.20 g, 86%) as a white needle: mp 179-181 °C; ³¹P NMR (C₅D₅N) δ 27.30; ¹H NMR (C₅D₅N) δ 0.92-2.18 (10H, m, 2,3,4,5,6-H of cyclohexylamine), 3.07 (1H, m, 1-H of cyclohexylamine), 7.34-7.44 (4H, 6,7-H of 2-naphthyl), 7.74-7.80 (6H, m, 4,5,8-H of 2-naphthyl), 8.18, (2H, d, J = 5.4 Hz, 3-H of 2-naphthyl), 8.49 (2H, s, 1-H of 2-naphthyl); ¹³C NMR (C₅D₅N) δ 22.27, 22.68, 29.04, 48.09, 121.47, 121.64, 125.95, 127.15, 129.20, 129.56, 129.61, 130.51, 130.87, 131.78. Anal. Calcd for C₂₆H₂₈NO₂PS₂: C, 64.84; H, 5.86; N, 2.91; S, 13.32. Found: C, 65.36; H, 6.09; N, 2.81; S, 13.03.

S,S'-Bis(1-naphthyl) 5'-O-[3-(Imidazol-1-ylmethyl)-4',4"-dimethoxy]tritylthymidine 3'-Phosphorodithioate (3c). 5'-O -[3-(Imidazol-1-ylmethyl)-4',4"dimethoxyltrityl-thymidine 1 (0.10 g, 0.16 mmol) and 2c (0.12 g, 0.24 mmol) were dried by repeated coevaporation with dry pyridine and dissolved in dry pyridine (1.6 ml). To the above solution was added isodurenedisulfonyl dichloride (0.15 g, 0.48 mmol). The mixture was stirred at rt for 30 min. The reaction was quenched by addition of 5% NaHCO₃ and the mixture was extracted three times with CHCl₃. The aqueous layer was back-extracted with CHCl3. The organic layer and washings were combined, dried over Na₂SO₄, filtered, and concentrated to dryness under reduced pressure. The residue was applied to a column of silica gel (4 g). Chromatography eluted with CHCl₃ containing 0.5% of methanol gave 3c (0.14 g, 90%) as a colorless foam: ^{31}P NMR (CDCl₃) δ 49.55; ¹H NMR (CDCl₃) δ 1.34 (3H, s, 5-CH₃), 1.99-2.03 (2H, m, 2',2"-H), 3.14 (2H, m, 5',5"-H), 3.70-3.76 (7H, m, 4'-H, OCH₃ of IDTr), 4.92 (2H, s, CH₂ of IDTr), 5.17-5.20 (1H, m, 3'-H), 6.07 (1H, dd J = 6.3 Hz, J = 7.3 Hz, 1'-H), 6.77-8.24 (31H, m, 6-H, ArH of IDTr, 1-naphthyl), 8.96 (1H, s, 3-H); ¹³C NMR (CDCl₃) δ 11.65, 24.94, 38.78, 38.85, 50.69, 55.22, 63.00, 77.20, 78.96, 79.08, 84.04, 84.35, 84.40, 86.87, 111.34, 113.39, 119.17, 122.73, 122.84, 122.95, 123.07, 125.61, 125.66, 125.77, 125.95, 126.69, 126.85, 127.37, 127.92, 128.46, 128.55, 128.66, 129.20, 129.78, 130.12, 130.98, 131.03, 131.09, 131.16, 134.23, 134.30, 134.54, 134.59, 134.63, 134.68, 135.06, 135.09, 135.98, 136.05,

136.14, 137.14, 145.09, 150.10, 158.83, 158.89, 163.45. Anal. Calcd for C₅₅H₄₉N₄O₈PS₂ •2H₂O: C, 64.44; H, 4.82; N, 5.47; S, 6.26. Found: C, 64.77; H, 5.20; N, 5.25; S, 7.01.

S,S'-Bis(2-naphthyl) 5'-*O*-[3-(Imidazol-1-ylmethyl)-4',4"-dimethoxy]tritylthymidine 3'-Phosphorodithioate (3d). The same procedure as described in the synthesis of 3c by use of 2d (0.12 g, 0.24 mmol) gave 3d (0.14 g, 90%) as a colorless foam: 31 P NMR (CDCl₃) δ 50.85; 1 H NMR (CDCl₃) δ 1.41 (3H, s, 5-CH₃), 2.25-2.36 (1H, m, 2'-H), 2.50-2.57 (1H, m, 2"-H), 3.30, 3.31 (2H, m, 5',5"-H), 3.76 (OCH₃ of IDTr), 4.17 (1H, m, 4'-H), 4.99 (2H, s, CH₂ of IDTr), 5.44-5.50 (1H, m, 3'-H), 6.40 (1H, dd J = 5.3 Hz, J = 8.8 Hz, 1'-H), 6.79-8.01 (31H, m, 6-H, ArH of IDTr, 2-naphthyl), 8.79 (1H, s, 3-H); 13 C NMR (CDCl₃) δ 8.48, 11.72, 24.82, 29.17, 33.89, 46.04, 51.23, 55.15, 62.91, 77.20, 84.34, 86.83, 111.32, 113.35, 119.59, 122.28, 122.59, 122.70, 126.24, 126.54, 126.92, 127.46, 127.69, 128.21, 128.73, 129.17, 129.24, 129.74, 130.08, 131.03, 131.09, 131.16, 131.21, 133.15, 133.19, 133.23, 133.33, 133.39, 133.44, 134.07, 134.95, 135.24, 135.35, 135.45, 135.54, 135.65, 136.44, 138.69, 145.30, 147.69, 149.02, 150.15, 158.80, 158.83, 153.49. Anal. Calcd for C₅₅H₄₉N₄O₈PS₂*2/3H₂O: C, 65.01; H, 4.86; N, 5.51; S, 6.31. Found: C, 65.13; H, 4.77; N, 5.31; S, 6.70.

Triethylammonium S-(2,6-Dichlorophenyl) 5'-O-[3-(Imidazol-1-ylmethyl)-4',4"-dimethoxy]tritylthymidine 3'-Phosphorothioate (4b). 5'-O -[3-(Imidazol-1vlmethyl)-4',4"-dimethoxyltritylthymidine 1 (0.10 g, 0.16 mmol) and 2c (0.12 g, 0.24 mmol) were dried by repeated coevaporation with dry pyridine and dissolved in dry pyridine (1.6 ml). To the above solution was added isodurenedisulfonyl dichloride (0.15 g. 0.48 mmol). The mixture was stirred at rt for 30 min. To the above solution, 2 M TEAB buffer (2 mL) was added. After being stirred at rt for 2 h, the mixture was diluted with CHCl3 and washed three times with 2 M TEAB buffer. The aqueous layer was back-extracted three times with CHCl₃. The organic layer and washings were combined, dried over Na₂SO₄, filtered, and concentrated to dryness under reduced pressure. The residue was applied to a column of silica gel (4 g). Chromatography eluted with CHCl3 containing 5% of methanol gave 4b (0.10 g, 65%) as a colorless foam: ³¹P NMR $(CDCl_3) \delta 11.97$; ¹H NMR $(CDCl_3) \delta 1.28 (9H, t, J = 7.3 Hz, CH_3 of Et_3N)$, 1.37 (3H, s, 5-CH₃), 2.29-2.72 (2H, m, 2', 2''-H), 3.01 (6H, q, J = 7.3 Hz, CH₂ of Et₃N), 3.30 (1H, m, 5'-H), 3.52 (1H, m, 5"-H), 3.77, 3.79 (6H, 2s, OCH₃ of IDTr), 4.47 (1H, m, 4'-H), 5.17 (2H, m, CH₂ of IDTr), 5.42 (1H, m, 3'-H), 6.39 (1H, dd J = 5.8 Hz, J = 7.3 Hz, 1'-H), 6.80-8.70 (20H, m, 6-H, ArH of IDTr and 2,6-dichlorophenyl), 8.70 (1H, bs, 3-H); ¹³C NMR (CDCl₃) δ 8.88, 11.50, 29.60, 39.84, 45.79, 50.77, 55.19, 64.24, 76.80, 77.21, 84.46, 85.19, 85.27, 86.79, 110.98, 113.32, 119.17, 125.70, 126.79, 128.10, 128.23, 128.46, 128.63, 129.20, 129.52, 129.78, 130.05, 130.30, 131.30, 131.32, 134.39, 135.65, 135.69, 136.26, 137.30, 141.56, 141.62, 145.28, 150.51, 158.67, 158.76, 163.79. Anal. Calcd for C₄₇H₅₄Cl₂N₅O₉PS: C, 58.38; H, 6.53; N, 7.24; S, 3.32. Found: C, 58.14; H, 5.98; N, 6.52; S, 4.14.

Triethylammonium S-(1-Naphthyl) 5'-O-[3-(Imidazol-1-ylmethyl)-4',4"-dimethoxy]tritylthymidine 3'-Phosphorothioate (4c). S,S'-bis(1-naphthyl) 5'-O-[3-(imidazol-1-ylmethyl)-4',4"-dimethoxy]tritylthymidine 3'-phosphorodithioate (3c). (0.069 g, 0.070 mmol) was dissolved in H₂O-Et₃N-pyridine (1:2:2, v/v/v, 5 mL) and the mixture was stirred at rt for 10 min. The mixture was diluted with CHCl₃ and washed three times with 2 M TEAB buffer. The aqueous layer was back-extracted three times with CHCl₃. The organic layer and washings were combined, dried over Na₂SO₄, filtered, and concentrated to dryness under reduced pressure to give 4c (0.067 g, quant) as a foam. This compound was used for the condensation with 3'-O-benzoylthymidine without further purification: ³¹P NMR (CDCl₃) δ 13.75.

Triethylammonium 5'-O-[3-(Imidazol-1-ylmethyl)-4',4"-dimethoxy]tritylthymidine 3'-[S-(2-Naphthyl)] Phosphorothioate (4d). The same procedure with 3d (0.129 g, 0.130 mmol) gave 4d (0.124 g, quant) as a foam: ^{31}P NMR (CDCl₃) δ 13.77.

Triethylammonium O-(2-Chlorophenyl) 5'-O-[3-(Imidazol-1-ylmethyl)-4',4"dimethoxyltritylthymidine 3'-Phosphorothioate (9). To a solution of 2-chlorophenyl phosphorodichloridate (0.07 g, 0.29 mmol) in dry THF (2.5 mL) were added 1H-1,2,4triazole (0.06 g, 0.86 mmol) and Et₃N (120 µL). The mixture was stirred at rt for 40 min. Compound 1 (0.12 g, 0.19 mmol, dried by repeated coevaporation with dry pyridine) in dry THF (2 mL) was added to the above mixture. After being stirred at rt for 2 h, the mixture was treated with hexamethyldisilathian (700 mL). The mixture was diluted with CHCl₃ and washed three times with 2 M TEAB buffer. The aqueous layer was back-extracted three times with CHCl₃. The organic layer and washings were combined, dried over Na₂SO₄, filtered, and concentrated to dryness. The residue was applied to a column of silica gel (3 g). Chromatography eluted with CHCl₃ containing 3% of methanol gave 9 (0.10 g, 58%) as a colorless foam: ${}^{31}P$ NMR (CDCl₃) δ 53.51, 54.27; ¹H NMR (CDCl₃) δ 1.28 (9H, t, J = 7.3 Hz, CH₃ of Et₃N), 1.36 (3H, s, 5-CH₃), 2.28-2.42 (1H, m, 2'-H), 2.61-2.72 (1H, m, 2"-H), 3.06 (6H, q, J = 7.3 Hz, CH₂ of Et₃N), 3.15-3.38 (1H, m, 5'-H), 3.48-3.66 (1H, m, 5"-H), 3.76, 3.78 (6H, 2s, OCH₃ of IDTr), 4.43 (1H, m, 4'-H), 5.18 (2H, m, CH₂ of IDTr), 5.63 (1H, m, 3'-H), 6..48 (1H, m, 1'-H), 6..69-7.63 (20H, m, 6-H, ArH of IDTr and 2-chlorophenyl), 9.98 (1H, bs, 3-H); ¹³C NMR (CDC1₃) δ 8.41, 10.77, 11.43, 22.75, 23.52, 28.70, 39.19, 45.64, 50.59, 50.64, 50.73, 53.34, 55.06, 63.95, 74.88, 84.22, 84.37, 84.57, 84.62, 85.34, 85.43, 86.38, 86.61, 86.81, 110.96, 113.17, 118.89, 119.16, 119.25, 121.85, 121.91, 121,94, 122.00, 123.83, 123.95, 125.25, 125.34, 125.45, 125.50, 125.61, 125.77, 126.47, 126.63, 127.06, 127.19,

127.37, 127.44, 128.10, 128.16, 128.45, 128.57, 128.66, 128.82, 129.09, 129.27, 129.38, 129.56, 129.76, 129.83, 129.90, 130.01, 130.26, 130.42, 130.71, 134.02, 134.14, 134.93, 135.40, 135.72, 135.81, 136.01, 136.12, 136.26, 137.05, 137.13, 145.01, 145.16, 146.61, 146.58, 148.57, 148.61, 148.72, 150.51, 150.58, 158.47, 158.51, 158.54, 158.63, 158.71, 163.81, 163.92.

3¹P NMR Study of Internucleotidic Bond Formation. A triethylammonium 5'-O-[3-(imidazol-1-ylmethyl)-4',4"-dimethoxy]tritylthymidine 3'-phosphorothioate derivative (4b-d or 9, 0.025 mmol) and 3'-O-benzoylthymidine (0.009 g, 0.025 mmol) were dried by repeated coevaporation with dry pyridine and dissolved in pyridine (250 μL). To the solution was added isodurenedisulfonyl dichloride (0.025 g, 0.075 mmol). After being stirred for 10 min, the mixture was quenched by addition of 5% NaHCO₃ (1 mL). The mixture was diluted with CHCl₃ and washed three times with 5% NaHCO₃. The aqueous layer was back-extracted three times with CHCl₃. The organic layer and washings were combined, dried over Na₂SO₄, filtered, and concentrated to dryness. The crude dimer (8b-d or 12) was dissolved in CDCl₃ (500 μL) and its ³¹P NMR spectrum was recorded. The ratio of the diastereomers was estimated on the basis of integration of the resonance signals.

Dithymidine Phosphorothioate (13). A crude mixture of the dimer 12 (0.049 g, 0.043 mmol) was treated with 1% TFA in CH₂Cl₂ (500 μ L) at rt for 30 min. To the mixture was added pyridine (100 μ L) and the solution was concentrated to dryness. The residue was dissolved in dioxane-H₂O (1:1, v/v, 500 μ L) and a mixture of 2-pyridinealdoxime (0.053 g, 0.43 mmol) and *N,N,N',N'*-tertramethylguanidine (51 μ L, 0.41 mmol) in dioxane-H₂O (1:1, v/v, 500 μ L) was added. After the mixture was stirred at rt for 24 h, 25% NH₃ was added and the mixture was stirred at rt for 24 h. The mixture was evaporated to dryness and dissolved in water. The solution was washed three times with ether and concentrated to a small volume. The crude product was purified by paper chromatography (Whatmann 3MM) eluted with 2-PrOH-25% NH₃-H₂O (7:1:2, v/v/v) to give dithymidine phosphorothioate 13 (302 A₂₆₀ units, 42%).

Enzymatic Characterization of the Major Isomer. The major isomer of dithymidine phosphorothioate 13 was separated by reversed phase HPLC. This isomer (0.5 A_{260} units) was treated with snake venom phosphodiesterase (4 μ L) in 0.5 M Tris•HCl buffer (pH 8.0, 40 μ L) at 37 °C for 3.5 h and the mixture was analyzed by reversed phase HPLC. In a similar manner, the major isomer (0.5 A260 units) was treated with nuclease P1 (10 μ L) in 50 mM sodium acetate buffer (pH 5.3, 50 μ L) at 37 °C for 3.5 h and the mixture was analyzed by reversed phase HPLC.

REFERENCES

- Zon, G. Pharm. Res. 1988, 5, 539-549. Uhlmann, E.; Peyman, A. Chem. Rev. 1990, 90, 543-584. Englisch, U.; Gauss, D. H. Angew. Chem. Int. Ed. Engl. 1991, 30, 613-629. Wagner, R. W. Nature 1994, 372, 333-335. Milligan, J. F.; Matteucci, M. D.; Martin, J. C. J. Med. Chem. 1993, 36, 1923-1937. Stein, C. A.; Cheng, Y.-C. Science 1993, 261, 1004-1021. Helene, C.; Toulme, J.-J. Biochem. Biophys. Acta 1990, 1049, 99-125.
- Zon, G.; Stec, W. J. In Oligonucleotides and Their Analogues: A Practical Approach; Eckstein, F., Ed.; IRL Press: Oxford University, 1991; pp 87-108.
 Zon, G., In Protocols for Oligonucleotides and Analogs, Methods in Molecular Biology, Vol. 20, Agrawar, S., Ed.; Humana Press: Totowa, New Jersey, 1993; pp 165-189.
- 3. Froehler, B. C. Tetrahedron Lett. 1986, 27, 5575-5578.
- 4. Susaka, A.; Grajkowski, A.; Wilk, A.; Uzanski, B.; Blaszczyk, J.; Wieczorek, M.; Stec, W. J. *Pure Appl. Chem.* **1993**, *65*, 707-714.
- 5. Lesnikowski, Z. J.; Sibinska, A. Tetrahedron 1986, 42, 5025-5034. Cosstick, R.; Williams, D. M. Nucleic Acids Res. 1987, 15, 9921-9932. FujiI, M.; Ozaki, K.; Kume, A.; Sekine, M.; Hata, T. Tetrahedron Lett. 1986, 26, 935-938. Fujii. M.; Ozaki, K.; Sekine, M.; Hata, T. Tetrahedron, 1987, 43, 3395-3407. Lesnikowski, Z. J.; Jaworska, M. Tetrahedron Lett. 1989, 30, 3821-3824. Seela, F.; Kretschmer, U. J. Chem. Soc., Chem. Commun. 1990, 1154-1159. Seela, F.; Kretschmer, U. J. Org. Chem. 1991, 56, 3861-3869. Stec, W. J.; Grajkowski, A.; Koziolkiewicz, M.; Uznanski, B. Nucleic Acids Res. 1991, 19, 5883-5888. Lesnikowski, Z. J. Nucleosides Nucleotides 1992, 11, 1621-1638. Stawinski, J.; Strömberg, R.; Zain, R. Tetrahedron Lett. 1992, 33, 3185-3188. Almer, H.; Stawinski, J.; Strömberg, R. *Nucleic Acids Res.* **1996**, 24, 3811-3820. Mizuguchi, M.; Kakino, K. Nucleosides Nucleotides 1996, 15, 407-417. Wang, J. C.; Just, G. Tetrahedron Lett. 1997, 38, 705-708. For a comprehensive review on the stereoselective synthesis of phosphorothioate oligonucleotides see Stec, W.; Wilk, A Angew. Chem. Int. Ed. Engl. 1994, 33, 709-722.
- 6. Sekine, M.; Hata, T. J. Org. Chem. 1987, 52, 946-948.
- 7. Sekine, M.; Mori, T.; Wada, T. Tetrahedron Lett. 1993, 34, 8289-8292.
- 8. Sekine, M.; Matsuzaki, J.; Hata, T.; Tetrahedron Lett. 1981, 22, 3209-3212.
- 9. Sekine, M.; Hamaoki, K.; Hata, T. J. Org. Chem. 1979, 44, 2325-2326.
- Garegg, P. J.; Regberg, T.; Stawinski, J.; Strömberg, R. Tetrahedron Lett.
 1986, 23, 2665-2668. Matsuzaki, J.; Hotoda, H.; Sekine, M.; Hata, T.
 Tetrahedron Lett. 1986, 25, 6545-2668.

11. Van Boom, J. H.; deRooy, J. F. M.; Reese, C. B. J. Chem. Soc. Perkin I, 1973, 2513-2517.

- 12. Wada, T.; Hata, T. Tetrahedron Lett. 1990, 31, 7461-7464.
- 13. Zain, R.; Strömberg, R.; Stawinski, J. J. Org. Chem. 1995, 60, 8241-8244.
- 14. Bryant, F. R.; Benkovic, S. J. Biochemistry, 1979, 18, 2825-2828.
- Potter, B. V. L.; Connolly, B. A.; Eckstein, F. *Biochemistry*, 1983, 22, 1369-1377.